

School/Department:	Erasmus Medical Center Department of Pathology
Project Title:	The role of endothelial-mesenchymal transition (EndMT) in human cancers
Abstract:	<p>The tumor microenvironment (TME) is the reactive cellular and humoral environment of a tumor (also designated as “tumor reactive stroma”). The TME includes the surrounding blood vessels, various cell types and components among which immune cells, fibrocytes, fibroblasts, myofibroblasts, the extracellular matrix (ECM) and the humoral milieu of signaling molecules, growth factors, chemokines and cytokines where these cells reside. The TME plays a critical role in tumor initiation and progression. The properties of the TME may limit the access of therapeutics to the tumor, may alter drug metabolism and may contribute to the development of drug resistance. As a consequence, the tumor microenvironment has a profound effect on tumor development, plasticity and response to therapy. The cellular and/or molecular stromal components of the TME are of paramount importance for biological significance and offer attractive therapeutic targets. The tumor and its surrounding microenvironment are closely related and interact constantly. The components of the TME are influenced by local as well as systemic factors. Tumor cells may adopt particular phenotypical and genotypical characteristics dependent on the environment in which the cells reside. Therefore, if tumor cells leave their microenvironment, or if the environment changes, the tumor cells may adopt completely new properties. Clearly, the TME is a complex environmental constellation with specific spatiotemporal and dynamic properties, which are defined by the in vivo situation and which cannot simply be reproduced in in vitro models. The population of fibroblasts in cancers (cancer-associated fibroblasts, CAF) is a prominent component of the TME and is supposed to have a profound effect on the tumor cells and other stromal cells in most types of solid tumors. The possible origins of CAFs include epithelial-mesenchymal transition (EMT), adipose-derived stem cells (ASCs) or carcinoma-associated adipocytes (CAAs), endothelial-mesenchymal transition (EndMT), bone marrow-derived cells and more. The endothelial-mesenchymal transition (EndMT) has largely been investigated in tissue fibrosis, not in cancers, and its role in human cancers remains largely unclear.</p> <p>In this PhD project, we will focus on the EndMT in human cancers and investigate the association of EndMT with motility and proliferation of the tumor cells, since cell migration and cell proliferation of cancer cells are the two key cellular phenomena for any types of cancer development and progression, irrespective of their molecular characteristics. In order to study this association with human relevance, human tumor tissues and/or xenograft animal models are required.</p> <p>The Department of Pathology of the Erasmus MC is equipped with a large arsenal of</p>

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	<p>techniques, including molecular methods for DNA, RNA, proteins, cyto-histologic analysis, PALM laser-capture microdissection systems, tissue culture facilities, and a variety of microscopes & imaging systems among which fluorescent microscopy, confocal laser scanning microscopy, (immune) electron microscopy, and more. There is a tissue bank in the department encompassing a large variety of tumor types. There are experiences with zebrafish modelling of development and diseases (collaboration with the Dept. of Clinical Genetics). A zebrafish model can be developed accordingly.</p>
Requirements of candidate:	<p>Master degree: Yes</p> <p>(Academic) Background: MD degree, preferably with interest in cancer biology and pathology. Strong motivation to obtain a PhD-degree within four years.</p> <p>IELTS Grade: 7.0 (<i>minimal 6.0 per component</i>) or TOEFL: 100 (<i>minimal 20 per component</i>)</p>
Supervisor information:	<p>Dr. P.P. Zheng (MD, PhD) / Prof. J.M. Kros (MD, PhD) Email address: p.zheng.1@erasmusmc.nl ; j.m.kros@erasmusmc.nl</p> <p>Selected publications:</p> <ol style="list-style-type: none"> 1. Kros JM, Mustafa DM, Dekker LJ, Sillevs Smitt PA, Luider TM and Zheng PP. Circulating glioma biomarkers. <i>Neuro Oncol</i>. 2014. DOI: 10.1093/neuonc/nou207 [Epub ahead of print] 2. Zheng PP, van der Weiden M and Kros JM. Fast tracking of co-localization of multiple markers by using the nanozoomer slide scanner and NDPViewer. <i>J Cell Physiol</i>. 2014;229:967-73. 3. Zheng PP, van der Weiden M, van der Spek PJ, Vincent AJ and Kros JM. Intratumoral, not circulating, endothelial progenitor cells share genetic aberrations with glial tumor cells. <i>J Cell Physiol</i>. 2013;228:1383-90. 4. Zheng PP, van der Weiden M and Kros JM. Video-Coupled Laser Capture Microdissection Using the PALM MicroBeam: A Powerful Approach for Live Digital Data Communications in Biomedical Research and Education. <i>J Mol Genet Med</i> 2013;7. DOI: 10.4172/1747-0862.1000069 5. Zheng PP, van der Weiden M, van der Spek PJ, Vincent AJ and Kros JM. Isocitrate dehydrogenase 1R132H mutation in microglia/macrophages in gliomas: indication of a significant role of microglia/macrophages in glial tumorigenesis. <i>Cancer Biol Ther</i>. 2012;13:836-9. 6. Zheng PP, van der Spek PJ, Dirven CM, Willemsen R and Kros JM. Sinus venosus defect (SVD) identified in zebrafish Glut1 morphants by video imaging. <i>Int J Cardiol</i>. 2012;154:e60-1. 7. Zheng PP, Romme E, van der Spek PJ, Dirven CM, Willemsen R and Kros JM. HeNe laser (633 nm)-coupled confocal microscope allows simulating magnetic resonance imaging/computed tomography scan of the brain and eye: a noninvasive optical approach applicable to small laboratory animals. <i>Zebrafish</i>. 2011;8:83-5. 8. Zheng PP, Romme E, van der Spek PJ, Dirven CM, Willemsen R and Kros JM. Defect of development of ocular vasculature in Glut1/SLC2A1 knockdown in vivo. <i>Cell Cycle</i>. 2011;10:1871 - 1872.

	<p>9. Zheng PP, Romme E, van der Spek PJ, Dirven CM, Willemsen R and Kros JM. Glut1/SLC2A1 is crucial for the development of the blood-brain barrier in vivo. <i>Ann Neurol</i>. 2010;68:835-44.</p> <p>10. Zheng PP, Severijnen LA, Willemsen R and Kros JM. Images. Different patterns of circulatory shunting in zebrafish caldesmon morphants: a digital motion analysis. <i>Heart Lung Circ</i>. 2010;19:251</p> <p>11. Zheng PP and Kros JM. Challenge of the gap between the current mania of cancer stem cells and the therapeutic strategy for patients with cancer. <i>Int J Cancer</i>. 2010;126:1529-30.</p> <p>12. Zheng PP, Severijnen LA, Willemsen R and Kros JM. Images in cardiovascular medicine. Functional cardiac phenotypes in zebrafish caldesmon morphants: a digital motion analysis. <i>Circulation</i>. 2009;120:e145-6.</p> <p>13. Zheng PP, Severijnen LA, Willemsen R and Kros JM. Circulation status of subintestinal vessels is a sensitive parameter for monitoring suboptimal systemic circulation in experimental zebrafish embryos. <i>Cell Cycle</i>. 2009;8:3782-3.</p> <p>14. Zheng PP, Severijnen LA, Willemsen R and Kros JM. Haemoglobin staining for in vivo portraying of functional vasculature in experimental zebrafish embryos. <i>Biochem Biophys Res Commun</i>. 2009;380:823-4.</p> <p>15. Zheng PP, Severijnen LA, van der Weiden M, Willemsen R and Kros JM. Cell proliferation and migration are mutually exclusive cellular phenomena in vivo: implications for cancer therapeutic strategies. <i>Cell Cycle</i>. 2009;8:950-1.</p> <p>16. Zheng PP, Severijnen LA, Willemsen R and Kros JM. Reduction of caldesmon expression induces apoptosis and causes disassembly of the sarcomeric protein complex in cardiomyocytes in vivo. <i>Cell Cycle</i>. 2009;8:325-6.</p> <p>17. Zheng PP, Severijnen LA, Willemsen R and Kros JM. Caldesmon is essential for cardiac morphogenesis and function: in vivo study using a zebrafish model. <i>Biochem Biophys Res Commun</i>. 2009;378:37-40.</p> <p>18. Zheng PP, Severijnen LA, van der Weiden M, Willemsen R and Kros JM. A crucial role of caldesmon in vascular development in vivo. <i>Cardiovasc Res</i>. 2009;81:362-9.</p> <p>19. Kros JM, van der Weiden M, Zheng PP, Hop WC, van den Bent MJ and Kouwenhoven MC. Intratumoral distribution of 1p loss in oligodendroglial tumors. <i>J Neuropathol Exp Neurol</i>. 2007;66:1118-23.</p> <p>20. Zheng PP, Weiden M, Sillevs Smitt PA, Luider TM and Kros JM. Hela +/-CaD undergoes a DNA replication-associated switch in localization from the cytoplasm to the nuclei of endothelial cells/endothelial progenitor cells in human tumor vasculature. <i>Cancer Biol Ther</i>. 2007;6:886-90.</p> <p>21. Kros JM, Gorlia T, Kouwenhoven MC, Zheng PP, Collins VP, Figarella-Branger D, Giangaspero F, Giannini C, Mokhtari K, Mork SJ, Paetau A, Reifenberger G and van den Bent MJ. Panel review of anaplastic oligodendroglioma from European Organization For Research and Treatment of Cancer Trial 26951: assessment of consensus in diagnosis, influence of 1p/19q loss, and correlations with outcome. <i>J Neuropathol Exp Neurol</i>. 2007;66:545-51.</p> <p>22. Zheng PP, Hop WC, Luider TM, Sillevs Smitt PA and Kros JM. Increased levels of circulating endothelial progenitor cells and circulating endothelial nitric oxide synthase in patients with gliomas. <i>Ann Neurol</i>. 2007;62:40-8.</p>
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	<p>23. Zheng PP, van der Weiden M and Kros JM. Hela l-CaD is implicated in the migration of endothelial cells/endothelial progenitor cells in human neoplasms. <i>Cell Adh Migr</i>. 2007;1:84-91.</p> <p>24. Zheng PP, Hop WC, Sillevs Smitt PA, van den Bent MJ, Avezaat CJ, Luider TM and Kros JM. Low-molecular weight caldesmon as a potential serum marker for glioma. <i>Clin Cancer Res</i>. 2005;11:4388-92.</p> <p>25. Zheng PP, van der Weiden M and Kros JM. Differential expression of Hela-type caldesmon in tumour neovascularization: a new marker of angiogenic endothelial cells. <i>J Pathol</i>. 2005;205:408-14.</p> <p>26. Zheng PP, Sieuwerts AM, Luider TM, van der Weiden M, Sillevs-Smitt PA and Kros JM. Differential expression of splicing variants of the human caldesmon gene (CALD1) in glioma neovascularization versus normal brain microvasculature. <i>Am J Pathol</i>. 2004;164:2217-28.</p> <p>27. Zheng PP, Luider TM, Pieters R, Avezaat CJ, van den Bent MJ, Sillevs Smitt PA and Kros JM. Identification of tumor-related proteins by proteomic analysis of cerebrospinal fluid from patients with primary brain tumors. <i>J Neuropathol Exp Neurol</i>. 2003;62:855-62.</p> <p>28. Zheng PP, Kros JM, Sillevs-Smitt PA and Luider TM. Proteomics in primary brain tumors. <i>Front Biosci</i>. 2003;8:d451-63.</p> <p>29. Kros JM, Bagdi EK, Zheng PP, Hop WC, Driesse MJ, Krenacs L and Dinjens WN. Analysis of immunoglobulin H gene rearrangement by polymerase chain reaction in primary central nervous system lymphoma. <i>J Neurosurg</i>. 2002;97:1390-6.</p> <p>30. Kros JM, Zheng PP, Dinjens WN and Alers JC. Genetic aberrations in gliomatosis cerebri support monoclonal tumorigenesis. <i>J Neuropathol Exp Neurol</i>. 2002;61:806-14.</p>
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